

Notification Date: November 4, 2022 Effective Date: December 6, 2022

# Inherited Motor and Sensory Neuropathy Gene Panel, Varies

Test ID: IMSNP

### **Useful for:**

- Establishing a molecular diagnosis for patients with hereditary motor and sensory neuropathy (HMSN) or Charcot-Marie-Tooth (CMT) disease
- Identifying variants within genes known to be associated with HMSN or CMT disease, allowing for predictive testing of at-risk family members

### **Genetics Information:**

- This test utilizes-next generation sequencing to detect single nucleotide and copy number variants in 87 genes associated with hereditary motor and sensory neuropathy: AARS1, ABCD1, AIFM1, ARSA, ATP1A1, ATP7A, BAG3, BSCL2, C1orf194, CHCHD10, CNTNAP1, COX10, COX6A1, CTDP1, DGAT2, DHH, DNM2, DYNC1H1, EGR2, ERCC8, FAM126A, FBLN5, FGD4, FIG4, FMR1, GALC, GAN, GARS1, GBF1, GDAP1, GJB1, GLA, GNB4, HARS1, HINT1, HK1, HSPB1, HSPB8, IGHMBP2, INF2, KARS1, LAMA2, LITAF, LMNA, LRSAM1, MARS1, MCM3AP, MFN2, MORC2, MPV17, MPZ, MTMR2, NDRG1, NEFH, NEFL, PEX7, PHYH, PLEKHG5, PLP1, PMP2, PMP22, PNKP, POLG, PRPS1, PRX, RAB7A, SACS, SBF1, SBF2, SCO2, SH3TC2, SLC12A6, SLC25A46, SORD, SOX10, SURF1, TDP1, TFG, TRIM2, TRPV4, TSFM, TTR, TUBB3, TWNK, TYMP, VPS13D, and YARS1. See Targeted Genes and Methodology Details for Inherited Motor and Sensory Neuropathy Gene Panel and Method Description for additional details.
- Identification of a disease-causing variant may assist with diagnosis, prognosis, clinical management, recurrence risk assessment, familial screening, and genetic counseling for hereditary motor and sensory neuropathy.

### Methods:

Sequence Capture and Targeted Next-Generation Sequencing followed by Polymerase Chain Reaction (PCR) and Sanger Sequencing

### **Reference Values:**

An interpretive report will be provided.

### **Specimen Requirements:**

**Patient Preparation:** A previous bone marrow transplant from an allogenic donor will interfere with testing. Call 800-533-1710 for instructions for testing patients who have received a bone marrow transplant.

Specimen Type: Whole blood

Preferred: Lavender top (EDTA) or yellow top (ACD)

Acceptable: Any anticoagulant

Specimen Volume: 3 mL

Collection Instructions: 1. Invert several times to mix blood

2. Send whole blood specimen in original tube. Do not aliquot.

Specimen Stability Information: Ambient (preferred)/Refrigerated

Minimum Volume: 1 mL

### Note:

Specimen preferred to arrive within 96 hours of collection.

# **Specimen Stability Information:**

Specimen Type	Temperature	Time	Special Container	
Varies	Varies			

# Ordering Guidance:

- First tier testing for a diagnosis of Charcot-Marie-Tooth disease type 1 is available; order PMPDD / PMP22 Gene, Large Deletion/Duplication Analysis, Varies.
- Targeted testing for familial variants (also called site-specific or known variants testing) is available for the genes on this panel. See FMTT / Familial Mutation, Targeted Testing, Varies. To obtain more information about this testing option, call 800-533-1710.
- Customization of this panel and single gene analysis for any gene present on this panel are available. For more information see CGPH / Custom Gene Panel, Hereditary, Next-Generation Sequencing, Varies.

#### Cautions:

#### **Clinical Correlations:**

- Test results should be interpreted in the context of clinical findings, family history, and other laboratory data. Misinterpretation of results may occur if the information provided is inaccurate or incomplete.
- If testing was performed because of a clinically significant family history, it is often useful to first test an
  affected family member. Detection of a reportable variant in an affected family member would allow for
  more informative testing of at-risk individuals.
- To discuss the availability of additional testing options or for assistance in the interpretation of these results, contact the Mayo Clinic Laboratories genetic counselors at 800-533-1710.

### **Technical Limitations:**

- Next-generation sequencing may not detect all types of genomic variants. In rare cases, false-negative or
  false-positive results may occur. The depth of coverage may be variable for some target regions; assay
  performance below the minimum acceptable criteria or for failed regions will be noted. Given these
  limitations, negative results do not rule out the diagnosis of a genetic disorder. If a specific clinical disorder
  is suspected, evaluation by alternative methods can be considered.
- There may be regions of genes that cannot be effectively evaluated by sequencing or deletion and duplication analysis as a result of technical limitations of the assay, including regions of homology, high

- guanine-cytosine (GC) content, and repetitive sequences. Confirmation of select reportable variants will be performed by alternate methodologies based on internal laboratory criteria.
- This test is validated to detect 95% of deletions up to 75 base pairs (bp) and insertions up to 47 bp. Deletions-insertions (delins) of 40 or more bp, including mobile element insertions, may be less reliably detected than smaller delins.

# **Deletion/Duplication Analysis:**

- This analysis targets single and multi-exon deletions/duplications; however, in some instances single exon
  resolution cannot be achieved due to isolated reduction in sequence coverage or inherent genomic
  complexity. Balanced structural rearrangements (such as translocations and inversions) may not be
  detected.
- This test is not designed to detect low levels of mosaicism or to differentiate between somatic and germline
  variants. If there is a possibility that any detected variant is somatic, additional testing may be necessary to
  clarify the significance of results.
- Genes may be added or removed based on updated clinical relevance. Refer to the Targeted Genes and Methodology Details for the Inherited Motor and Sensory Neuropathy Gene Panel for the most up to date list of genes included in this test. For detailed information regarding gene specific performance and technical limitations, see Method Description or contact a laboratory genetic counselor.
- If the patient has had an allogeneic hematopoietic stem cell transplant or a recent blood transfusion, results may be inaccurate due to the presence of donor DNA. Call Mayo Clinic Laboratories for instructions for testing patients who have received a bone marrow transplant.

### **Reclassification of Variants:**

At this time, it is not standard practice for the laboratory to systematically review previously classified
variants on a regular basis. The laboratory encourages healthcare providers to contact the laboratory at
any time to learn how the classification of a particular variant may have changed over time. Due to
broadening genetic knowledge, it is possible that the laboratory may discover new information of relevance
to the patient. Should that occur, the laboratory may issue an amended report.

### **Variant Evaluation:**

- Evaluation and categorization of variants are performed using published American College of Medical
  Genetics and Genomics and the Association for Molecular Pathology recommendations as a guideline.(1)
  Other gene-specific guidelines may also be considered. Variants are classified based on known, predicted,
  or possible pathogenicity and reported with interpretive comments detailing their potential or known
  significance. Variants classified as benign or likely benign are not reported.
- Multiple in silico evaluation tools may be used to assist in the interpretation of these results. The accuracy
  of predictions made by in silico evaluation tools is highly dependent upon the data available for a given
  gene, and periodic updates to these tools may cause predictions to change over time. Results from in silico
  evaluation tools should be interpreted with caution and professional clinical judgment.
- Rarely, incidental or secondary findings may implicate another predisposition or presence of active disease. These findings will be carefully reviewed to determine whether they will be reported.

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Day(s) Performed: Varies Report Available: 28 to 42 days

### Questions

Contact Michelle Raths, Laboratory Resource Coordinator at 800-533-1710.